

Competition Between Weeds and Pepper in Southern Italy

Stella Lovelli*, Teodoro Di Tommaso, Mariana Amato, Maria Valerio, Michele Perniola

Crop System, Forestry and Environmental Sciences Department, University of Basilicata
Viale dell'Ateneo Lucano 10, 85100 Potenza, Italy

Received: 14 September 2009. Accepted: 12 January 2010.

Abstract

In arid areas drought conditions and warmer temperatures will alter the competitive balance between crops and some weed species. The objective of this study was to study water competition and its effect on canopy relationship of a C4 weed (pigweed) and a C3 weed (bindweed) towards a C3 crop (pepper) in a Mediterranean area. The experiment was carried out in 2008 in Matera, Southern Italy. Pigweed and bindweed were studied within a naturally occurring weed population in a bell pepper field where a rainfed treatment (V0) was compared to a full irrigated one (V100), the latter corresponding to the restoration of 100% of the maximum crop evapotranspiration, (ET_c). Soil water content was measured periodically; leaf water potential, net assimilation rate (A), stomatal conductance (gs), transpiration rate (T), Ci (intercellular CO₂ concentration) and A/Ci curves were also determined on pigweed, bindweed and pepper leaves.

All gas exchange parameters differed between irrigated and rainfed treatments and between the three species. Water use efficiency was higher in pigweed than in pepper and bindweed. Between the considered weeds, pigweed competed for water with pepper significantly since, unlike bindweed, pigweed began to reduce stomatal conductance only when its leaf water potential achieved very negative values, lower than -2.00 MPa. Unlike C4 crops already saturated for CO₂, pigweed photosynthesis is not completely saturated for CO₂. Consequently, since atmospheric CO₂ is increasing, when pigweed is grown in mixed stands where competition occurs, it can further limit other slow-growing species, both crops and weeds.

Key-words: Pigweed, *Amaranthus retroflexus* L., Bindweed, *Convolvulus arvensis* L., leaf water potential, stomatal conductance, water use efficiency.

Introduction

Some authors (Bajaj et al., 1999) estimated that two-thirds of the potential yield of major crops are usually lost due to adverse growing environments. Moreover, currently weed control is one of the greatest costs for farmers. As known weeds interfere with crop growth and depress yield by competing for available resources (Zimdahl, 2004). In addition, future growing conditions for crops will be altered by climate change in several ways, including greater atmospheric [CO₂], higher temperature, altered timing and quantity of water availability, greater tropospheric [O₃] and altered incidence of pests, diseases and pollinator actions. In addition, most

climate change scenarios predict a worldwide increase of arid areas throughout the world, including the Mediterranean basin (IPPC, 2001 and 2007).

In natural ecosystems where species with the C3 and C4 pathways contemporarily occur, the periods of their most active growth are temporally separated so that competition is little intense (Patterson, 1995). In the agroecosystem, the growth and competitive interactions of C3 and C4 crops and weeds are synchronized by the timing of crop sowing and by other management practices (Patterson, 1995).

In these last years with the certainty that Earth's climate is changing and that global warming seems to be inevitable, regardless of

* Corresponding Author: Tel.: +39 0971 205384; Fax: +39 0971 205378. E-mail address: stella.lovelli@unibas.it

future gas emission reductions, it has become very important to identify potential vulnerabilities and adaptive responses in managed ecosystems (Howden et al., 2007).

Some authors (Tungate et al., 2007) underlined that warmer temperatures will alter the competitive balance between crops and weed species, intensifying weed pressures. Competition for water by weeds and effect of weed growth on yield is well recognized (French and Schultz, 1984b). Notwithstanding, field-based information on weed physiology is essential to achieve a better understanding of how changes in temperature may alter weed growth and competition relationships with crops.

Pigweed (*Amaranthus retroflexus* L.) is a summer annual invasive plant, widely distributed in Italy and in other areas of the world in cultivated lands. It is difficult to manage in agronomic crops because of prolonged germination period, relatively fast growth, high seed production and long-term seed viability. Some authors (Horak and Loughin, 2000, Lovelli et al., 2010) observed that pigweed is an increasingly aggressive weed in semiarid environments and that, in general, the control of *Amaranthus* species is becoming very difficult.

Field bindweed (*Convolvulus arvensis* L.) is a perennial weed in Europe and many agricultural areas of the world (Weaver and Riley, 1982). This weed has been described as the twelfth worst weed in the world (Holm et al., 1977). In agroecosystem bindweed reduces crop value through competition and by interfering with the harvest procedures (Pfirter et al., 1997). Moreover, field bindweed provides a breeding site for insects attacking adjacent crops (Tamaiki et al., 1975). In some cases it serves as an alternative host for viruses which cause plant diseases (Holm et al., 1977). Bindweed control by both mechanical and chemical methods is very difficult because of its high regeneration ability

(Pfirter et al., 1997). It is possible to stop shoot growth and reduce the amount of root by repeating herbicide applications but, even after several years applications, the growth of some roots, which can allow further shoot development, remains (Timmons, 1949).

The objective of this study was to study water competition of a C4 weed (pigweed) and a C3 weed (bindweed) with a C3 crop (bell pepper) at canopy level, in a Mediterranean area.

Materials and methods

Plant material and growth conditions

The experiment was carried out in 2008 in Matera, Southern Italy (40°00' N; 16°00' E; 397 m a.s.l.) on a loam soil with a moderate chemical fertility. Soil moisture content was 24.2% at field capacity and 17.2% at the theoretical wilting point (determined in lab at -0.03 and -1.5 MPa respectively). Plots consisted of six 30 m rows, spaced 1 m apart. Soil tillage consisted of moldboard ploughing, disking and land levelling previous to crop planting. Bell pepper (*Capiscum annuum* L.) (cv. peppone) was transplanted on 19-05-08, 1 m x 0.4 m spaced and gradually harvested until 26/08/2008. During the trial 150 kg ha⁻¹ of N, 150 kg ha⁻¹ of P₂O₅ and 180 kg ha⁻¹ of K₂O were applied. Naturally occurring weed populations were used in the experiments (Tab. 1).

Experimental design

Two different irrigation treatments, arranged in the field as a split-plot design with four replications, were compared: rainfed (V0) and fully irrigated (V100), where 100% of total crop evapotranspiration (ET_c) was restored. Irrigation was applied when 40% of total available water was depleted according to the evapotranspirometric method of Doorenbos and Pruitt

Table 1. Densities of weeds observed on field at harvesting of pepper.

Latin name	Common name	Density (plants m ⁻²)	
		V0	V100
<i>Amaranthus retroflexus</i> L.	Pigweed	3 ± 0.2	30 ± 0.2
<i>Convolvulus arvensis</i> L.	Bindweed	1 ± 0.1	15 ± 3.0
<i>Chenopodium album</i> L.	Lamsquarters	3 ± 0.2	6 ± 1.2
<i>Portulaca oleracea</i> L.	Purslane	22 ± 0.6	12 ± 4.5

Values are means ± standard error.

(1977). ET_c was calculated as $ET_c = ET_o \times K_c$ where ET_o (reference evapotranspiration) was calculated according to Hargreaves and Samani (1985):

$$ET_o = 0.0023 (T_{\text{mean}} + 17.8) (T_{\text{max}} - T_{\text{min}})^{0.5} R_a \quad [1]$$

where: T_{mean} , T_{max} , T_{min} mean, maximum and minimum air temperature in °C, respectively and R_a is the extraterrestrial solar radiation in mm d^{-1} (tabulated value). K_c was the crop coefficient of bell pepper as reported by Allen et al., (1998), corrected for the specific environmental conditions, yielding $K_{c \text{ ini}} = 0.5$; $K_{c \text{ med}} = 1.15$; $K_{c \text{ end}} = 0.8$. Drip irrigation was used, with dripping wings placed on each row and “on line” drippers, spacing 20 cm, with a 3 l h^{-1} delivery. Photosynthetic Active Radiation, max and min T , air humidity and wind speed were acquired every 10 min by a weather station placed in a meadow next to the plots, averaged and recorded every 30 min by a datalogger (Model Sky DataHog2, type SDL5400).

Soil water status

Soil water content was periodically measured during the experiment and at the moment of photosynthetic measurements by gravimetric method at 0-30 cm and 30-60 cm depth and expressed as soil water potential, after determining the soil water retention curve in the lab.

Plant water status

Leaf water potential (Ψ) was measured on pigweed, bindweed and pepper plants 40 days after transplantation of bell pepper. Measurements were made on the youngest uppermost fully-expanded leaf of four representative plants per treatment, using pressure chamber technique (Scholander et al., 1965).

Gas exchange analysis

Instantaneous gas exchange measurements were made on pigweed, bindweed and pepper plants 40 days after transplantation of the main crop on the youngest uppermost fully-expanded leaf of four representative plants per treatment. Net assimilation rate (A), stomatal conductance (g_s), transpiration rate (T) and C_i (intercellular CO_2 concentration) were simultaneously determined, using the youngest, fully expanded 40

days old leaf. These measurements were made on four plants for each treatments. A portable open – gas exchange system (Model LiCor-6400, Li Corporation, Lincoln, NE, USA) was used, incorporating infrared CO_2 and water vapour analyzers and using a saturating red light source at a PPFD (Photosynthetic Photon Flux Density) of $1800 \mu\text{mol m}^{-2}\text{s}^{-1}$. The cuvette temperature was held at the mean air temperature at noon on the measurement day (33°C) and the relative humidity within the cuvette was maintained at 20%. Water use efficiency at the leaf level (WUE_l) was calculated as the ratio between assimilated CO_2 and transpired H_2O ($\mu\text{mol CO}_2 \text{ mmol}^{-1} \text{ H}_2\text{O}$).

The response of A to C_i was assessed only on irrigated plots (V100) by changing the concentration of CO_2 entering the leaf chamber (O_2 was maintained at $210 \text{ mmol mol}^{-1}$). Three plants assayed over three consecutive days were used per treatment. Each sample consisted of one fully-expanded leaf of pigweed, bindweed and pepper plants from each plot. Measurements for the A/C_i response curve were carried out with a saturating red light source at a PPFD of $1800 \mu\text{mol m}^{-2} \text{ s}^{-1}$ and maintaining leaf temperature near to the air temperature in field on that day ($33\text{--}34^\circ\text{C}$). The CO_2 concentration of the cuvette was initially maintained at $380 \mu\text{mol mol}^{-1}$ for 5 min to stimulate stomatal opening, then reduced to 300, 200, 100, 0 and then increased to 100, 200, 300, 400, 500, 700, 800, 900, 1200, 2000 $\mu\text{mol mol}^{-1}$ and gas exchange properties were logged after the system had achieved a predetermined stability point. Curve fitting software was used to analyze the A/C_i response curves using the following three component exponential function:

$$A = a(1 - e^{-bx}) + c \quad [2]$$

where A = steady-state assimilation rate and $x = C_i$ (intercellular CO_2 concentration).

In interpreting the CO_2 response of photosynthesis in pepper and pigweed (C3 plants) carboxylation efficiency (K) was calculated as the initial slope of the curve by the following equation:

$$K = A/(C_c - \Gamma) \quad [3]$$

by Farquhar and Sharkey 1982, where A is the net assimilation rate at atmospheric CO_2 , C_c is the CO_2 concentration inside the chloroplast

(assumed to be equal to C_i) and Γ is the compensation point. By equation [2] the A_{\max} (maximum assimilation rate) was calculated as $a + c$.

In interpreting the CO_2 response of photosynthesis in pigweed, the model of C4 photosynthesis developed by Von Caemmerer and Furbank (1999) was used. By equation [2], the A_{sat} (CO_2 saturated rate) was calculated as $a + c$. This last value is an indicator of the ribulose-1,5-bisphosphate carboxylase/oxygenase activity (Rubisco activity), of the Phosphoenolpyruvate regeneration rate and electronic transport rate. The CE value, indicator of Phosphoenolpyruvate carboxylase (PEPC) activity was calculated as the slope at $A = 0$ (calculated as $b[a + c]$).

Statistical analysis

The statistical design was a split-plot with four replications. Analysis of variance (ANOVA) was performed and mean discrimination was carried out according to the Duncan's Multiple Range Test. Significant differences were accepted at $P < 0.05$ and $P < 0.01$ and represented by different letters. To compare A/C_i curves we combined all the data sets to obtain a unique curve (combine model). Parameters of this unique curve were then compared with the specific model of each curve with the F-test according to the following equation:

$$F = \frac{\frac{RSS_c - \sum RSS_i}{df_c - \sum df_i}}{\frac{\sum RSS_i}{\sum df_i}} \quad (3)$$

where RSS_c = residual sum of square of the combine model; RSS_i = residual sum of square of each curve; df_c = residual degrees of freedom of the combine model; df_i = residual degrees of freedom of each curve. All the analysis were carried out using the statistical package Sigma Plot 11.0 for Windows (USA).

Results

In the irrigated treatment the most present weeds at pepper harvesting were pigweed (*Amaranthus retroflexus* L., C4 plant) and bindweed (*Convolvulus arvensis* L., C3 plant). Other weeds also observed on the bell pepper

Table 2. Leaf water potential of pepper, pigweed and bindweed measured 40 days after pepper transplantation.

	Leaf water potential (MPa)	
	V0	V100
Pepper	-1,7 ± 0.09	-1,1 ± 0.07
Pigweed	-2,4 ± 0.04	-1,7 ± 0.04
Bindweed	-1,9 ± 0.08	-1,4 ± 0.13

Values are means ± standard error.

field were purslane (*Portulaca oleracea* L.) and common lambsquarters (*Chenopodium album* L.). In irrigated treatment pigweed prevailed, while in drought conditions purslane prevailed (Tab. 1).

A significant decrease in soil water potential was observed in rainfed conditions. At the moment of photosynthetic measurements, 40 days after transplantation of the main crop, the average soil water potential for the whole 0-60 cm depth soil volume was -0.96 MPa in V100 plots and -2.24 MPa in V0 plots. The average soil water status of -2.24 MPa reported in the V0 treatment corresponds to 0.74 MPa below the theoretical wilting point. Consequently, highly significant differences in plant water status were found at noon among treatments (Tab. 2). The lower value was measured on pigweed in rainfed treatment, while the higher leaf water potential was measured on irrigated pepper, -2.4 and -1.1 MPa, respectively (Tab. 2).

All gas exchange parameters differed between irrigated and rainfed treatments and among the three species (Tab. 3). We observed a significant effect of drought conditions on photosynthetic rate of all three species. Of course, higher value of net assimilation, was recorded in pigweed, 37.6 $\mu\text{mol m}^{-2}\text{s}^{-1}$ and 13.9 $\mu\text{mol m}^{-2}\text{s}^{-1}$ in V100 treatment and in V0 treatment, respectively (Tab. 3). Lower assimilation values were measured in bindweed, 3.0 and 1.1 $\mu\text{mol m}^{-2}\text{s}^{-1}$ in V100 treatment and in V0 treatment, respectively. Transpiration rate was higher in V100 treatment than in V0 treatment; stomatal conductance was significantly lower in rainfed conditions in the three species. Comparing pepper and pigweed we observed that, in drought treatment, in correspondence to the same stomatal conductance and transpiration values, in pigweed photosynthesis is nearly three times as much as pepper. Consequently, water use efficiency was higher in pigweed than in

Table 3. Gas exchange parameters of pepper, pigweed and bindweed leaves in irrigated (V100) and not irrigated (V0) conditions.

	A			T			gs			WUE		
	V0	V100	mean	V0	V100	mean	V0	V100	mean	V0	V100	mean
Pepper	4.9 BC	12.0 BC	8.5	3.4	5.6	4.5 AB	0.07C	0.26 A	0.16	1.4	2.2	1.8 B
Pigweed	13.9 B	37.6 A	25.6	3.9	7.8	5.8 A	0.08 C	0.30 A	0.19	3.5	4.8	4.2 A
Bindweed	1.1 C	3.0 BC	2.0	1.7	4.4	3.1 B	0.06 C	0.20 B	0.13	0.7	0.8	0.7 B
mean	6.6	17.5		3.0 B	5.9 A		0.07	0.25		1.9	2.5	

Values within a column followed by different letters are significantly different at $P \leq 0.01$ according to Duncan's Multiple Range Test. Number of replicates = 4.

pepper on average, 3.5 and 1.4 $\mu\text{mol}/\text{m}^2 \text{mol H}_2\text{O}$, respectively.

The relationship between stomatal water vapour conductance and leaf water potential is also reported (Fig. 1). Our data show that stomatal conductance in all three species decreases as leaf water status achieves low values. But stomatal conductance begins first decreasing in pepper, when leaf water potential is lower than -1.25 MPa, then in bindweed at lower than -1.0 MPa water potential, and only at very negative value of leaf water potential in pigweed, lower than -2.00 MPa.

From A/Ci curves emerges that pepper and bindweed show the classic C3 plants response while pigweed has a typical C4 plant behaviour (Fig. 2). Examination of A/Ci curves indicates that K (initial slope), maximum photosynthetic

rate (asymptote), and photosynthetic rate measured at atmospheric CO_2 concentration were different in pepper and bindweed. Carboxylation efficiency (K) was higher in pepper than in bindweed, 0.26 ± 0.03 and $0.14 \pm 0.01 \text{ molm}^{-2}\text{s}^{-1}$, respectively. Also, maximum photosynthetic rate (asymptote) was higher in pepper than in bindweed, as statistical analysis highlighted ($P < 0.05$).

In pigweed (C4 plant) our data show a greater photosynthetic activity of pigweed than pepper and bindweed, as expected. PEPC activity (CE) was $0.6 \mu\text{molm}^{-2}\text{s}^{-1}$, while A_{sat} was $84.9 \mu\text{mol m}^{-2}\text{s}^{-1}$, as we reported in a previous paper (Lovelli et al., 2010). Moreover our data show that, for pigweed in the Mediterranean environment, photosynthesis is not completely saturated for CO_2 . In fact, as figure 2 clearly shows,

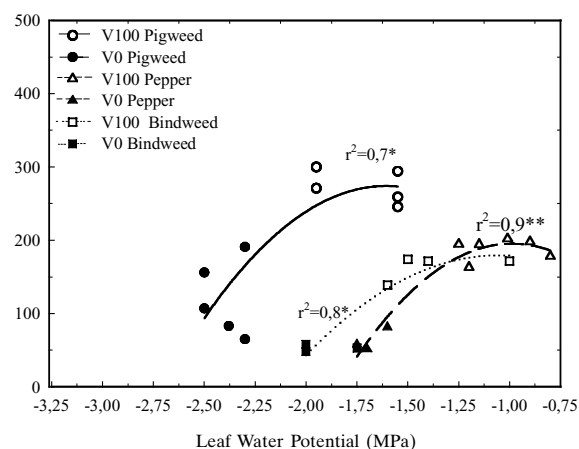


Figure 1. Stomatal water vapour conductance as a function of leaf water potential measured on pepper, pigweed and bindweed leaves in irrigated treatment (V100, open symbols, all data measured) and not irrigated treatment (V0, closed symbols, all data measured). Data were fit using non linear regression analysis. Lines are polynomial fit for all data points.

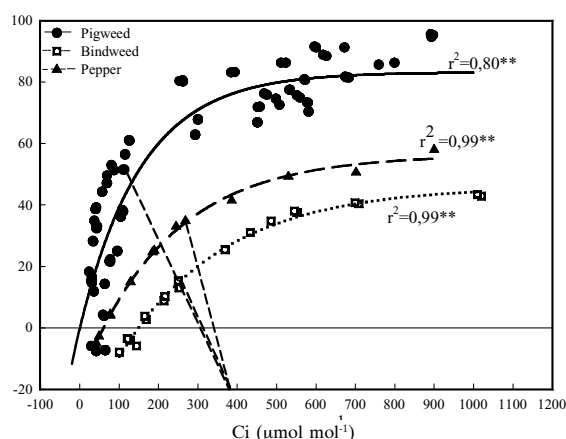


Figure 2. Response of leaf photosynthesis (measured as leaf CO_2 assimilation rate, A) to a range of internal CO_2 concentration (C_i) for single leaves of pepper, pigweed and bindweed, only in irrigated plots (V100, all points obtained from four replicates). Lines are exponential fit for all data points. The dashed lines from the x axis indicate the CO_2 assimilation rate at atmospheric CO_2 concentration (operating C_i).

in pigweed the operating C_i of photosynthesis under ambient CO_2 concentration is below the inflexion point of the A/C_i curve.

Discussion

We observed that photosynthetic rate decreases as stomatal CO_2 conductance decreases both in pepper and weeds. Our data show that stomatal conductance decreases as leaf water status achieves low values, as expected. Although there were other weeds in each plot (Tab. 1) if we compare pigweed and bindweed only, the first one significantly competes with pepper for water, since unlike bindweed, pigweed begins reducing stomatal conductance only when its leaf water potential achieves very negative values, lower than -2.00 MPa. In this sense, pigweed showed to be more drought resistant than pepper and bindweed. From a physiological point of view, both C3 and C4 species do not differ in the extent to which they resist severe drought, and the main difference in the performance as a response to drought conditions is a greater water use efficiency of the C4 pathway (Osmond et al., 1982). At the operating C_i in normal air for non-stressed plants, C4 species have over twice the WUE of C3 plants with an equal assimilation rate (Osmond et al., 1982; Knapp and Medina, 1999). In this way C4 plants can reach the same CO_2 assimilation rate as C3 plants with about half the stomatal conductance, and thus half the rate of water loss (Sage and Kubien, 2003). Hence for a given amount of soil water, C4 plants can develop a larger canopy, grow more root mass and produce more seeds than their C3 competitors (Ludlow, 1985; Long, 1999). A larger leaf canopy has other effects since it can shade slower growing C3 species, while a larger root system gives to C4 plants the possibility to uptake soil resources before C3 competitors (Sage and Kubien, 2003).

The response of photosynthesis to intercellular CO_2 is frequently used to evaluate the mechanisms controlling photosynthetic responses to environmental change, in both C3 and C4 species (Sharkey, 1985). Analysis of A/C_i response curves shows that the CO_2 assimilation rate increases as ambient CO_2 increases in C3 plants, as expected, but also in pigweed, a C4 plant. Actually, in the latter species the operat-

ing C_i of photosynthesis under ambient CO_2 concentration is below the inflexion point of the A/C_i curve. Other authors (Ghannoum et al., 2000; Leakey, 2009) also concluded that in the future C4 photosynthesis could only be directly stimulated by elevated CO_2 concentration when the operating C_i of photosynthesis under ambient CO_2 is below the inflexion point of the A/C_i curve. This is an important result that could affect competition and increase weed aggressiveness towards crops in agro-ecosystems. As well known, the direct physiological effects of increased atmospheric CO_2 probably will be most beneficial to C3 plants (Patterson, 1995). However, the likely climatic consequences of the global warming, i.e. increased temperature and aridity, are most likely to favour C4 plants, and between them, C4 weeds, such as pigweed. It will be possible not only because of the well-known C4 pathway superiority in drought condition but also because CO_2 unsaturated weeds will remain sensitive to higher ambient CO_2 levels (Sage and Kubien, 2003). Thus, when they are grown in mixed stands where competition occurs, the fast-growing C4 species can control the slow-growing species (Grise, 1996), especially C3 crops.

Acknowledgment

This research was funded by CLIMESCO Italian Project, contract N. 286 of 20/Feb/2006 (Ministry of University and Research).

References

- Allen R.G., L.S. Pereira, D. Raes, M. Smith 1998. Crop evapotranspiration. Guidelines for computing crop water requirements. FAO Irrig. Drain. Paper N. 56, FAO, Rome, Italy.
- Bajaj S., Targolli J., Liu L.F., Ho T.H.D., Wu R. 1999. Transgenic approaches to increase dehydration-stress tolerance in plants. *Mol. Breed.*, 5:493-503.
- Doorenbos J., Pruitt W.O. 1977. Crop water requirements. FAO Irrig. Drain. Paper N. 24. FAO, Rome, Italy, 144 pp.
- Farquhar G.D., Sharkey T.D. 1982. Stomatal conductance and photosynthesis. *Annual Review of Plant Physiology*, 33:317-345.
- French R.J., Schultz J.E. 1984b. Water use efficiency of wheat in a mediterranean-type environment. II. Some limitations to efficiency. *Australian Journal of Agricultural Research*, 35:765-775.

- Ghannoum O., von Caemmerer S., Ziska L.H., Conroy J.P. 2000. The growth response of C4 plants to rising atmospheric CO₂ partial pressure: a reassessment. *Plant Cell Environ.*, 23:931-942.
- Grise D.J. 1996. Effects of elevated CO₂ and high temperature on the relative growth rates and competitive interactions between a C3 (*Chenopodium album*) and a C4 (*Amaranthus hybridus*) annual. PhD thesis, University of Georgia, Athens, Georgia.
- Hargreaves G.H., Samani Z.A. 1985. Reference Crop evapotranspiration from temperature. *Applied Eng. Agric.*, 1:96-99.
- Holm L.G., Plucknett D.L., Pancho J.V., Herberger J.P. 1977. *The World's Worst Weeds: Distribution and Biology*. Honolulu: University Press of Hawaii.
- Horak M.J., Loughin T.M. 2000. Growth analysis of four *Amaranthus* species. *Weed Sci.*, 48:347-355.
- Howden S.M., Soussana J.F., Tubiello F.N., Chhetri N., Dunlop M., Meinke H. 2007. Adapting agriculture to climate change. *PNAS* 104, 19691-19696.
- Intergovernmental Panel on Climatic Change (IPCC), 2001. In: Houghton J.T., Ding Y., Griggs D.J., Noguer M., van Der Linden P.J., Xiasou D. (eds.): *The Scientific Basis. Third Assessment Report of Working Group I*. Cambridge University Press, Cambridge.
- Intergovernmental Panel on Climatic Change (IPCC), 2007. *Fourth Assessment Report of Working Group I. Climate Change 2007: The Physical Science Basis, Summary for Policymakers*.
- Knapp A.K., Medina E. 1999. Success of C4 photosynthesis in the field: Lessons from communities dominated by C4 plants. In: Sage R.F., Monson R.K. (eds.): *C4 Plant Biology*, 251-283. Academic Press, San Diego, California.
- Leakey A.D.B. 2009. Rising atmospheric carbon dioxide concentration and the future of C4 crops for food and fuel. *Proceedings Royal Society Botany*, 276:2333-2343. doi:10.1098/rspb.2008.1517.
- Long S.P. 1999. Environmental responses. In: Sage R.F., Monson R.K. (eds.): *The Biology of C4 Photosynthesis*, 215-249. Academic Press, San Diego, California.
- Lovelli S., Perniola M., Ferrara A., Amato M., Di Tommaso T. 2010. Photosynthetic response to water stress of pigweed (*Amaranthus retroflexus* L.) in a Southern-Mediterranean area. *Weed Science*, 58:126-131.
- Ludlow M.M. 1985. Photosynthesis and dry matter production in C3 and C4 pasture plants, with special emphasis on tropical C3 legumes and grasses. *Aust. J. Plant Physiol.*, 12:557-572.
- Osmond C.B., Winter K., Ziegler H. 1982. Functional significance of different pathways of CO₂ fixation in photosynthesis. In: Lange O.L., Nobel P.S., Osmond C.B., Ziegler H. (eds.): *Encyclopedia of Plant Physiology, New Series, Vol. 12B. Physiological Plant Ecology II. Water Relations and Carbon Assimilation*, 479-547. Springer-Verlag, Berlin.
- Patterson D.T. 1995a. Weeds in a changing climate. *Weed Science*, 43:685-701.
- Pfister H.A., Ammon H.U., Guntli D., Greaves M.P., De Fago G. 1997. Towards the management of field bindweed (*Convolvulus arvensis*) and hedge bindweed (*Calystegia sepium*) with fungal pathogens and cover crops. *Integrated Pest Management Reviews*, 2:61-69.
- Sage R., Kubien D. 2003. Quo vadis C4? An ecophysiological perspective on global change and the future of C4 plants. *Photosynth. Res.*, 77: 209-225.
- Sharkey T.D. 1985. Photosynthesis in intact leaves of C3 plants. Its occurrence and a possible explanation. *Botanical Review*, 51:53-105.
- Scholander P.F., Hammel H.T., Bradstreet E.D., Hemmingsen E.A. 1965. Sap pressure in vascular plants. *Science*, 148:339-346.
- Tamaki G., Moffitt H.R., Turner J.E. 1975 The influence of perennial weeds on the abundance of the red-backed cutworm on asparagi. *Environmental Entomology*, 4:274-276.
- Timmons F.L. 1949. Duration of viability of bindweed seed under field conditions and experimental results in the control of bindweed seedlings. *Agronomy Journal*, 1949:130-133.
- Tungate K.D., Israel D.W., Watson D.M., Rufty T.W. 2007. Potential changes in weed competitiveness in an agroecological system with elevated temperatures. *Environ. Exp. Bot.*, 60:42-49.
- von Caemmerer S., Furbank R.T. 1999. Modeling C4 photosynthesis. IL: R.F. Sage, R.K. Monson, eds, *C4 Plant Biology*. Academic Press, San Diego, 173-211.
- Weaver S.E., Riley W.R. 1982. The biology of Canadian weeds. 53. *Convolvulus arvensis*. *Canadian Journal of Plant Science*, 62:461-72.
- Zimdahl R.L. 2004. *Weed-crop competition: A review*. Ames, IA: Blackwell Publishing, 27-106.

